PII: S0040-4020(96)00899-X

SYNTHETIC STUDY ON CURACIN A: A NOVEL ANTIMITOTIC AGENT FROM THE CYANOBACTERIUM Lyngbya majuscula

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Abstract: Curacin A (1), a novel antimitotic agent, was synthesized in a highly stereo-controlled manner. The four stereoisomers of a partial structure at the thiazoline moiety, 2 were also synthesized to aid in elucidation of the absolute configurations of three chiral centers in curacin A. The effects on porcine brain tubulin assembly of several synthetic compounds related to curacin A were examined.

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Microtubules are the main component of spindles in the mitotic apparatus of eucaryotic cells, and are also involved in other cell functions, such as axonal transport, motility, and determination of cell shape. The major constituent of the microtubule system is the protein tubulin.\(^1\) There are a number of natural and synthetic compounds that interfere with tubulin function to inhibit the formation of microtubules and to cause the mitotic arrest of eucaryotic cells. Such antimitotic agents show a broad spectrum of biological activities, and have potential applications in the fields of medicine and agriculture. They can also be used as molecular probes for investigating the dynamics of microtubule networks. Studies on the mechanism of molecular recognition between inhibitors and tubulin are important, because the inhibitor-binding domain is expected to have a close relationship to the domain that functions in tubulin assembly.\(^2\)

Curacin A (1) is a novel antimitotic agent isolated from a Caribbean cyanobacterium, Lyngbya majuscula,³ and consists of a disubstituted thiazoline bearing a chiral cyclopropane ring and an aliphatic side chain. It was reported that curacin A inhibits tubulin assembly by binding to the colchicine-binding site³, which is one of the two distinct drug-binding sites on tubulin. This result is intriguing because curacin A has little structural similarity to known natural and synthetic colchicine-site ligands. Thus, elucidation of the nature of curacin A-binding to tubulin should afford further insight into the molecular mechanism of tubulin-ligand interaction at this site, and could lead to the development of new bioactive agents.

Several groups have reported synthetic approaches to curacin A.⁴⁻⁶ Here we describe a highly stereo-controlled total synthesis of curacin A and synthesis of the four stereoisomers of a partial structure of curacin A, 2-(2-methyl)cyclopropyl-4-(1-propenyl)thiazoline, (+)-2a, (+)-2b, (-)-2a and (-)-2b, aiming at the elucidation of the absolute configuration at positions 2, 19 and 21 of curacin A.^{5, 6d} We also discuss the structure-activity relationships of curacin A, and related molecules.

In the first paper on the structure of curacin A^3 , the E, E geometry of the conjugated diene (C(7-10)) and e relative configuration at the cyclopropyl moiety (C(19) and (21)) were established. Subsequently, the absolute structure of curacin A was determined to be 2R, 13R, 19R and 21S by chemical degradation and total synthesis by White et at. In the course of our study on curacin A, we first planned to synthesize the four stereoisomers of 2-(2-methyl)cyclopropyl-4-(1-propenyl)thiazoline, (+)-2a, (+)-2b, (-)-2a and (-)-2b, since one of them should compose the partial structure of curacin A and was expected to be a significant structural element in the interaction with tubulin. We intended to define the absolute configuration at the three chiral centers of the thiazoline-methylcyclopropane moiety in curacin A by comparison of the physicochemical data of 2a and 2b with those of curacin A, as well as from their biological activities. In our synthesis of the thiazolines 2, we used a selective deprotection of the N, S-acetal 5 to give the N-Boc amino thiol 6, which was readily converted to the thiol ester 8. The thiazoline ring was constructed by a facile conversion of 8 via the thiol ester ammonium salt 9.

The starting material, 3, was easily prepared from L-cysteine hydrochloride in three steps. 8,9 Reduction of the amide 3 with LiAlH₄ gave the corresponding aldehyde 4 in 92% yield. Wittig reaction of 4 with ethyltriphenylphosphonium bromide gave a mixture of olefins (Z/E=11/1). The Z/E ratio of 5 was determined from the 1 H-NMR integration of the H-5 methylene proton peak of the thiazoline moiety at δ 3.26 ppm. The major Z-isomer 5 could be purified by recrystallization from aqueous MeOH in 56% yield (Z/E=56/1).9 In deprotection of the N, S-acetal group of 5, it was found that 5 was unstable on exposure to TFA in CH₂Cl₂ at high concentrations or for long times, and the addition of thiophenol or ethanethiol was ineffective. However, selective deprotection of the N, S-acetal group proceeded in diluted TFA in water-saturated CH₂Cl₂ to give the N-Boc amino thiol 6 in 64% yield. The thiol 6 was converted to the thiol ester 8 using (±)-cis-2methylcyclopropanecarboxylic acid 7¹⁰ and bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BOPCl)¹¹. Deprotection of the tert-Boc group of 8 with TFA gave the thiol ester ammonium salt 9, which was heated to reflux in benzene to yield the thiazolines 2a and 2b as a mixture of diastereomers (42% yield from 6) containing traces of the E-isomer. The double bond geometry of each olefin isomer was assigned based on the ¹H-NMR coupling constants of 10.7 Hz (desired Z-isomer) and 15.1 Hz (E-isomer) for the olefinic protons. The mixture of diastereomers was found to be easily separable by HPLC using a TSK-gel, ODS-120T column to give (+)-2a ($|\alpha|_{l_{0}}^{25}$ +141.6° (c 0.94, CHCl₁)) and (+)-2b ($|\alpha|_{l_{0}}^{25}$ +108.4° (c 1.02, CHCl₁)). The corresponding (4S)-isomers were synthesized by the same procedure from D-cysteine hydrochloride to give (-)-2a ($[\alpha]_{D}^{25}$ -139.9° (c 0.90, CHCl₃)) and (-)-2b ($[\alpha]_{D}^{25}$ -104.3° (c 1.04, CHCl₃)), respectively (Scheme 1).

Scheme 1

Reagents and conditions: (a) LiAlH₄, ether, 0°C, 0.5 h (92%); (b) Ph₃PC₂H₅Br, n-BuLi, THF, 0°C, 0.5 h, then **4**, THF, 5°C, 2.5 h, and recrystallization (56%, Z/E=56/1); (c) TFA, CH₂Cl₂, 20°C, 2 h (64%); (d) (\pm) -cis-2-methylcyclopropanecarboxylic acid (\pm) -7, BOPCI, Et₃N, CH₂Cl₂, 20°C, 5 h; (e) TFA, CH₂Cl₂, 0°C, 1.5 h; (f) benzene, reflux, 1 h (42% from **6**)

In order to identify the stereochemistry of the four isomers thus prepared, an authentic sample of (+)-2a ($[\alpha]_D^{25}$ +145.6° (c 1.025, CHCl₃)) was synthesized from the *N*-Boc amino thiol 6 and (+)-(1S, 2R)-2-methylcyclopropanecarboxylic acid ((+)-7) (Scheme 2). The acid (+)-7 was prepared by optical resolution of the (±)-7 quinine salt.¹⁰ Recrystallization of (±)-7 quinine salt from acetone four times gave pure colorless crystals (mp 138-140.5 °C, $[\alpha]_D^{25}$ -125.0° (c 0.515, CHCl₃)) and treatment of the crystals in aqueous HCl regenerated the desired carboxylic acid ((+)-7) ($[\alpha]_D^{25}$ +24.9° (c 0.495, 95% EtOH)). The configurations of the other three isomers, (+)-2b, (-)-2a and (-)-2b, could accordingly be assigned from their ¹H-NMR spectra and optical rotations.

Scheme 2

Reagents: (a) quinine, acetone, crystallization, 4 times, then HCl, aq. EtOH; (b) $\bf 6$, BOPCl, Et₃N, CH₂Cl₂; (c) TFA, CH₂Cl₂; (d) benzene, reflux (34%)

¹H-NMR data of curacin A compared with those of the isomers of 2 showed that the chemical shifts of curacin A were not greatly different from those of (+)-2a and (+)-2b, but that the values of the coupling constants were closer to those of (+)-2b (Figure 1 and Table 1), suggesting that the relative stereochemistry of curacin A at positions 2, 19 and 21 is the same as that of 2b. The relatively large optical dextrorotation of (+)-2b ($[\alpha]_{\rm D}^{25}$ +108.4° (c 1.02, CHCl₃)) was compatible with that of curacin A ($[\alpha]_{\rm D}^{20}$ +62.0° (c 1.10, CHCl₃))^{6b}. Based on these physicochemical data, the absolute configuration of the partial structure of curacin A was concluded to be 2R, 19R and 21S. Our result is in agreement with that reported independently by White *et al.*^{4,6a}

Figure 1. H-NMR chemical shifts of (+)-2a, (+)-2b and curacin A

Table 1. ¹H-NMR coupling constants of (+)-2a, (+)-2b and curacin A

Compound	J _{5a, 5b} (Hz)	J _{5a, 4} (Hz)	J _{5b, 4} (Hz)
(+)-2a	10.8	6.7	8.4
(+)-2b	10.8	9.8	8.3
Curacin A ^{a)}	10.7	10.0	_ b)

a) Reported by Gerwick et al. 3 b) Not available

Contrary to our expectation, none of the four isomers of 2 showed inhibitory activity either on microtubule assembly, even at $100~\mu M$ concentration under the conditions used¹², or on some cancer-derived cell lines, even at $100~\mu g/mL$ concentration. Therefore, the approach to confirm the stereochemistry of curacin A by evalution of the biological activities of 2 was not feasible.

Next, we planned an asymmetric total

synthesis of curacin A (1). The retrosynthetic analysis and disconnections are shown in **Scheme 3**. We expected that the necessary three double bond-geometries could be prepared from geraniol (C(9-10)) by

semistabilized Wittig or Wittig-Horner reaction (C(7-8)) and Wittig reaction (C(3-4)). The chiral centers at C(2) and C(13) should be derived from a chiral synthon (L-cysteine) and an asymmetric allylation using a chiral allylationium reagent¹³, respectively. The chiral methylcyclopropane moiety could be efficiently prepared from diethyl L-tartrate, using a double-asymmetric Simmons-Smith cyclopropanation as a key step.¹⁴ We intended to construct the thiazoline moiety by coupling of the carboxylic acid (-)-7 with the N-Boc thiazolidine 29 through selective deprotection of the N, S-acetal group, using a similar procedure to that employed for 2. Since the chiral center attached to the nitrogen atom readily epimerizes and the thiazoline ring is unstable under basic conditions¹⁵, this ring system should be constructed at the final stage of the total synthesis.

Scheme 3

In regioselective epoxidation of geraniol using the OXONE®-acetone system¹⁶, the 2, 3:6, 7-diepoxide was obtained in 70% yield at high conversions. At moderate conversions of geraniol, however, production of the 2, 3:6, 7-diepoxide was considerably reduced and the desired 6, 7-epoxide 10 was obtained in 39% total yield (two cycle, plus 37% recovery of geraniol). Simultaneous acid-catalyzed hydrolysis and acetalization gave the 1, 3-dioxolane 11, a synthetic equivalent of aldehyde. First, we intended to form the conjugated E, E-diene unit by Wittig reaction of 12 with 15. Compound 11 was converted, *via* the bromide¹⁷, to the corresponding phosphonium salt 12 in 69% yield. Monoprotection¹⁸ of 1, 4-butanediol with p-methoxybenzyl chloride gave the PMB-protected alcohol 14 in 84% yield, and this was oxidized to the PMB-protected aldehyde 15 in 89% yield by Swern's method. Wittig reaction of 12 with 15 (n-BuLi, THF) afforded the desired diene 13 in 73% yield with low E selectivity (E/Z=1.9/1). The E/Z ratio of 13 was determined from the ¹H-NMR integration of the olefinic protons at H-6 (the E-isomer, δ 5.83 ppm; the Z-isomer, δ 6.12 ppm). The mixture of olefins was found to be separable by HPLC using a packed silica gel column to give the desired E-isomer (**Scheme 4**). In this Wittig reaction, the influence of a number of factors, such as solvent

(ether, CH₂Cl₂, toluene and DME), base (KHMDS, NaHMDS and t-BuOK), inorganic halides¹⁹ (LiI), excess reactant, reaction time and temperature, on the reaction stereochemistry was investigated. However, all attempts to improve the stereoselectivity were unsuccessful.

Reagents and conditions: (a) OXONE®, acetone-CH₂Cl₂/phosphate buffer, pH 7.5-8.0, 0°C, 2 h (39% and recovery of geraniol, 37%); (b) PTSA, aq. acetone, 20°C, 2 h (67%); (c) CBr₄, Ph₃P, CH₂Cl₂, 0°C, 1 h; (d) PPh₃, benzene, 20°C, 16 h (69% from 11); (e) 12, n-BuLi, THF, -78°C, 0.5 h, then 15, THF, -78-20°C, 16 h (73%, E/Z=1.9/1), and HPLC separation; (f) PMBCI, NaH, DMF, -15-20°C, 1.5 h (84%); (g) DMSO, (COCI)₂, Et₃N, CH₂Cl₂, -50-20°C, 1.5 h (89%)

In order to obtain the diene 13 with higher E selectivity, Wittig-Horner reaction²⁰ of the corresponding phosphonate 16 with 15 was employed. Compound 11 was converted, via the bromide, to the phosphonate 16 in 75% yield. Wittig-Horner reaction of 16 with the aldehyde 15 afforded the diene 13 (51%, E/Z=8.5/1) (Scheme 5). In this reaction, the desired diene could be obtained only when t-BuOK was added to a mixture of 16 and 15 in THF. Since the aldehyde 15 was unstable under this reaction condition, the use of excess amounts of 15 improved the yield.

Scheme 5

Reagents and conditions: (a) CBr₄, Ph₃P, CH₂Cl₂, 0°C, 1 h; (b) (EtO)₃P, benzene, reflux, 2.5 h (75% from 11); (c) 15, t-BuOK, THF, 20°C, 1.5 h (51%, E/Z=8.5/1 and recovery of 16, 13%)

The side chain of 1 was finally synthesized as shown in **Scheme 6**. Deacetalization of 13 with PTSA (two cycle) followed by oxidative cleavage of the diol 17 (three cycle) gave the aldehyde 18 in 93% total yield. The asymmetric allylation of 18 with a chiral allylation reagent¹³, prepared from [(4R, 5R)-2, 4R]

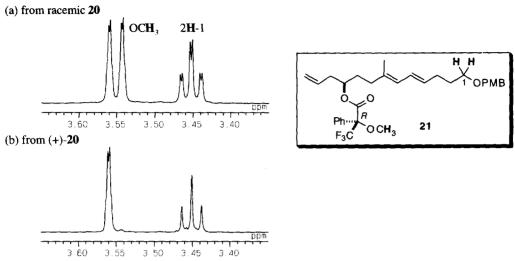
2-dimethyl-1, 3-dioxolane-4, 5-bis(diphenylmethoxy)]cyclopentadienyl-chlorotitanium ((R, R)-19) and allylmagnesium chloride, proceeded cleanly at -78°C to give the homoallylic alcohol 20 in 95% yield and with excellent enantioselectivity (>99% ee), as determined from the ¹H- and ¹³C-NMR spectra of its Mosher ester 21 (Figure 2).

Scheme 6

Reagents and conditions: (a) PTSA, aq. MeOH, 20° C, 5 h (99%); (b) NaIO₄, aq. acetone, 20° C, 2 h (94%); (c) allyIMgCl, (*R*, *R*)-19, THF, 0° C, 1 h, then 18, THF, -78°C, 1.5 h (95%); (d) (*S*)-(+)-MTPACl, pyridine, CH₂Cl₂, 20° C, 0.5 h (66%); (e) MeI, NaH, DMF, 20° C, 2.5 h (89%); (f) MgBr₂·OEt₂, Me₂S, CH₂Cl₂, 20° C, 2 h (76% and recovery of 22, 6%); (g) MsCl, pyridine, 0° C, 1 h, then NaI, acetone, reflux, 2 h (87%); (h)Ph₃P, MeCN, reflux, 7 h (quant.); (i) 25, LiHMDS, THF, -78°C, 0.5 h, then MeCHO, THF, -78-0°C, 1.5 h (41%)

The alcohol **20** was converted to its methyl ether **22** in 89% yield. In deprotection of the PMB group in **22**, treatment with DDQ or CAN resulted a complex mixture, but treatment with TFA in CH₂Cl₂²¹ gave a mixture of the desired alcohol **23** and decomposition products. We next sought a suitable condition to cleave the PMB ether in **22** by using a variety of combinations of hard Lewis acids and soft nucleophiles²², and found that MgBr₂·OEt₂-Me₂S treatment in CH₂Cl₂ was optimal to give the known and desired alcohol (-)-**23** in 76% total yield (five cycle).^{68, 23} In ether, this system resulted in complete recovery of the starting material, and the use of ethanethiol instead of Me₂S gave a mixture containing **23** and ethanethiol-adducts. The alcohol **23** was converted, *via* the iodide **24**, to the phosphonium salt **25** according to the reported procedure^{6a}. In order to obtain information on the structure-activity relationships of curacin A, the tetraene **26**, a lipid side chain moiety of curacin A, was also prepared through Wittig reaction of **25** with acetaldehyde in 41% yield (**Scheme 6**).





Asymmetric synthesis of the cyclopropane moiety of 1 is shown in Scheme 7. We have developed a versatile method for the synthesis of enantiomerically pure *cis*-2-methylcyclopropanecarboxylic acid 7.¹⁴ Double-asymmetric Simmons-Smith cyclopropanation of the diene 27 derived from diethyl L-tartrate in 4 steps proceeded with excellent diasterofacial selectivity (>99% de) to give the dicyclopropane 28, which was converted to the desired carboxylic acid (-)-7 in 3 steps.

Scheme 7

EtO₂C CO₂Et
Diethyl L-tartrate

27

Double-asymmetric Simmons-Smith cyclopropanation

28

$$\frac{3 \text{ steps}}{CO_2H} = \frac{[\alpha]_0^* - 32.6^{\circ}}{(c \ 0.495, 95\% \text{EtOH})}$$
(-)-(1*R*, 2*S*)-7

The total synthesis of curacin A was completed as shown in **Scheme 8**. Wittig reaction of the phosphonium salt **25** with the aldehyde **4** afforded the thiazolidine **29** in 60% yield. None of the *E*-isomer was detected by 1 H-NMR analysis. The ylide generated from **25** with LiHMDS was very sensitive to reaction temperature and moisture. Therefore the phosphonium salt **25** was azeotropically dehydrated by refluxing in dry benzene for 1 h just before use, and a liquid N₂-ethanol cooling bath was used to keep the temperature of the reaction mixture at < -78 ${}^{\circ}$ C during additions of LiHMDS and the aldehyde **4**. The thiazoline moiety of **1** was synthesized from the *N*-Boc thiazolidine **29** in a stepwise manner. Selective deprotection of the *N*, S-acetal group of **29** was carried out in diluted TFA in water-saturated CH₂Cl₂ to give the *N*-Boc amino thiol

30 (three cycle, 54% total yield), which was converted to the corresponding thiol ester using the carboxylic acid (-)-7 and bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BOPCl). Deprotection of the *tert*-Boc group of the thiol ester, followed by refluxing in benzene, gave curacin A in 10% yield from **29**. The physicochemical properties (¹H- and ¹³C-NMR spectra, optical rotation) of the synthesized curacin A are identical to those reported.^{3, 6b}

Scheme 8

Reagents and conditions: (a) **25**, LiHMDS, THF, -78°C, 0.5 h, then **4**, THF, -78-0°C, 2 h (60% and recovery of **4**, 25%); (b) TFA, CH_2CI_2 , 20°C, 6 h; (c) (-)-7, BOPCI, Et_3N , CH_2CI_2 , 20°C, 3 h; (d) TFA, CH_2CI_2 , 20°C, 2 h, then benzene, reflux, 2.5 h (10% from **29**)

The effects of the synthesized curacin A and the related compounds, 2, 22, 23, 26 and 29, on microtubule assembly were examined. Curacin A showed high anti-tubulin activity ($IC_{50}=2.5 \mu M$) under the conditions used¹², whereas the PMB ether 22, the alcohol 23, the tetraene 26, the *N*-Boc thiazolidine 29 and all the diastereomers of the thiazoline 2 did not inhibit tubulin polymerization. These results demonstrate that the combination of heterocyclic and lipid side chain moieties in curacin A is important for its anti-tubulin activity.

In conclusion, we propose the absolute configurations of three chiral centers in curacin A to be 2R, 19R and 21S, based on the spectral and physical data of four stereoisomers, (+)-2a, (+)-2b, (-)-2a and (-)-2b, synthesized as a partial structure of curacin A. We also achieved the total synthesis of curacin A in a highly stereo-controlled manner. Further studies on the structure-activity relationship of curacin A are in progress.

Acknowledgement

This work was supported in part by Grants-in-Aid for Scientific Research (No. 07772087 and 08672414) from the Ministry of Education, Science and Culture, Japan. We are grateful to Dr. Rudolf O. Duthaler, Ciba-Geigy AG, Basle, for his generous gift of the amide 3 and for valuable information. We also thank Dr. Naoko Morisaki for FABMS and HRFABMS measurements, Mrs. Hiroko Hino for elemental analyses and Dr. Kazuo Furihata for his valuable advice on NMR measurements.

Experimental Section

All ¹H- and ¹³C-NMR spectra were measured in CDCl₃ with TMS and the solvent peak as internal standards, and recorded on a JEOL JMN-A500 spectrometer. IR spectra were recorded on a JASCO A-102

infrared spectrophotometer. Mass spectra (MS) were obtained on a JEOL JMS-HX110 spectrometer. Optical rotations were measured on a JASCO DIP-100 digital polarimeter. All reactions were carried out in an atmosphere of dry argon at room temperature unless otherwise stated. Column chromatography was carried out on Wakogel C-200. Analytical thin-layer chromatography (TLC) was performed on Merck precoated silica gel 60 F_{254} plates, and compounds were visualized by UV illumination (254 nm) or by heating to 150 °C after spraying phosphomolybdic acid in ethanol. Dry diethyl ether (Et₂O) and tetrahydrofuran (THF) were distilled from sodium-benzophenone ketyl and dry benzene (PhH) and dichloromethane (CH₂Cl₂) were distilled from phosphorus pentoxide and calcium hydride, respectively under an inert atmosphere. Anhydrous N, N-dimethylformamide (DMF) and OXONE® (containing potassium peroxomonosulfate, KHSO₅) were purchased from Aldrich. [(4R, 5R)-2, 2-Dimethyl-1, 3-dioxolane-4, 5-bis(diphenylmethoxy)]cyclopentadienyl-chlorotitanium, (R, R)-19 was purchased from Fluka AG. All other organic solvents and reagents were obtained from commercial sources and used without further purification. Organic extracts were dried over magnesium sulfate (MgSO₄), filtered, and concentrated using a rotary evaporator at < 40 °C bath temperature. Involatile oils and solids were vacuum dried at < 2 mmHg.

(4R)-N-(tert-Butyloxy)carbonyl-2, 2-dimethyl-4-(N-methoxy-N-methylcarbamoyl)thiazolidine (3).

The title compound was prepared according to the method of Duthaler⁹: mp 97.0-97.5 °C, lit. mp 99.0-99.5 °C⁶; $[\alpha]_D^{25}$ -65.3° (c 1.02, CHCl₃), lit. $[\alpha]_D^{20}$ -64.1° (c 1.25, CHCl₃)⁹; IR (CHCl₃) 3000, 2950, 1700, 1680, 1460, 1390, 1370, 1320, 1170, 1130, 1090, 1070, 1010, 860 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃, 2 rotamer) δ 1.41 and 1.49 (s, 9H, (CH₃)₃C), 1.78, 1.83, 1.89 and 1.91 (s, 6H, (CH₃)₂-2), 2.99 (dd, 1H, J=12.0, 3.6 Hz, H-5), 3.22 (s, 3H, NCH₃), 3.34 (dd, 1H, J=12.0, 7.3 Hz, H-5), 3.73 and 3.78 (s, 3H, NOCH₃), 5.04 and 5.17 (m, 1H, H-4); FABMS m/z: 305 (M+H)⁺; HRFABMS calcd. for $C_{13}H_{25}O_4N_2S$ (M+H)⁺ 305.1535, found 305.1546.

(4S)-enantiomer was prepared according to the method above: mp 98.0-98.5 °C; $[\alpha]_{\rm b}^{25}$ +62.4° (c 1.02, CHCl₃)

(4R)-N-(tert-Butyloxy)carbonyl-2, 2-dimethyl-4-formylthiazolidine (4).

LiAlH₄ (2.18 g, 57.4 mmol) was added portionwise to a solution of 3 (25.0 g, 82.1 mmol) in dry Et₂O (300 mL) at 0 °C. The reaction mixture was stirred vigorously for 0.5 h at 0 °C and diluted with Et₂O (300 mL). After the slow addition of aqueous KHSO₄ (5.0 g, 36.7 mmol/H₂O 15 mL), the resulting slurry was filtered through Celite and the precipitate was washed well with Et₂O. The combined filtrates were washed with aqueous HCl (0.1 N, 200 mL), saturated aqueous NaHCO₃ and NaCl (200 mL), dried and concentrated. Kugelrohr distillation of the residue (150-180 °C, 1.5 mmHg) gave 4 (18.54 g, 92% yield) as a colorless oil: $[\alpha]_D^{25}$ -97.1° (*c* 1.01, CHCl₃), lit. $[\alpha]_D^{20}$ -99.1° (*c* 0.94, CHCl₃)⁹; IR (CHCl₃) 3000, 2950, 1750, 1710, 1680, 1460, 1400, 1370, 1290, 1170, 1130, 1100, 1080, 860 cm⁻¹; ¹H-NMR (500 MHz, C₆D₆, 70 °C) δ 1.34 (s, 9H, (CH₃)₃C), 1.70 and 1.79 (s, 6H, (CH₃)₂-2), 2.57 (dd, 1H, J=12.0, 6.6 Hz, H-5), 2.61 (dd, 1H, J=12.0, 3.0 Hz, H-5), 4.30 (br, 1H, H-4), 9.38 (s, 1H, CHO); FABMS m/z: 246 (M+H)⁺; HRFABMS calcd. for C₁₁H₂₀O₃NS (M+H)⁺ 246.1164, found 246.1170.

(4S)-enantiomer was prepared according to the method above: $[\alpha]_{\rm p}^{25}$ +98.3° (c 1.03, CHCl₃)

(4R)-N-(tert-Butyloxy)carbonyl-2, 2-dimethyl-4-[(1Z)-1-propen-1-yl]thiazolidine (5).

A suspension of ethyltriphenylphosphonium bromide (37.6 g, 0.101 mol) in dry THF (370 mL) was cooled to 0 °C and n-BuLi (58.6 mL, 1.6 M in hexane, 93.8 mmol) was added dropwise to the suspension. The reaction mixture was stirred for 0.5 h at 0 °C. A solution of 4 (18.41 g, 75.0 mmol) in dry THF (74 mL) was added dropwise to the resulting dark red solution at 0 °C over 20 min. The reaction mixture was stirred for 2.5 h at 5 °C and quenched slowly by the addition of a mixed solution of aqueous NaOH (1 M, 110 mL) and NaH₂PO₄ (1 M, 190 mL). The resulting mixture was filtered and the filtrate was extracted with EtOAc (2x220 mL). The combined organic phases were dried and concentrated to afford a white insoluble solid, which was washed with hexane/EtOAc (10/1, 350 mL). The washing solution was concentrated and

chromatography of the residue on silica gel (645 g, 30/1 hexane/EtOAc) gave **5** (16.74 g, 87% yield, E/Z=10.7/1) as a colorless solid. Recrystallization of the solid from aqueous MeOH (2.7/1 MeOH/H₂O, 110 mL) at 0 °C gave **5** (10.77 g, 56% yield, E/Z=55.9/1) as pure clear, colorless crystals: mp 33.0-33.5 °C, lit. mp 34.0 °C°; $[\alpha]_D^{25}$ +46.7° (c 0.985, CHCl₃), lit. $[\alpha]_D^{20}$ +48.9° (c 0.45, CHCl₃), R (CHCl₃) 3000, 2950, 1690, 1480, 1380, 1340, 1300, 1190, 1120, 1090, 1000, 940, 860 cm⁻¹; H-NMR (500 MHz, CDCl₃) δ 1.45 (s 9H, (CH₃)₃C), 1.71 (dd, 3H, J=6.9, 1.5 Hz, CH₃-2'), 1.78 and 1.80 (s, 6H, (CH₃)₂-2), 2.58(dd, 1H, J=11.6, 2.1 Hz, H-5), 3.26 (dd, 1H, J=11.6, 6.2 Hz, H-5), 5.11 (br, 1H, H-4), 5.50 (dq, 1H, J=10.8, 6.9 Hz, H-2'), 5.68 (ddq, 1H, J=10.8, 8.9, 1.5 Hz, H-1'); 13 C-NMR (125 MHz, CDCl₃) δ 13.12, 28.50 (x3), 29.65, 30.13, 34.06, 60.79, 70.25, 79.91, 124.44, 132.11, 152.46; FABMS m/z: 258 (M+H)+; HRFABMS calcd. for C₁₃H₂₄O₂NS (M+H)+ 258.1528, found 258.1524; Anal. Calcd for C₁₃H₂₃O₂NS: C, 60.66; H, 9.01; N, 5.44; S, 12.46. Found: C, 60.84; H, 9.05; N, 5.39; S, 12.28.

(4*S*)-enantiomer was prepared according to the method above: mp 31.5-32.0 °C; $[\alpha]_{\rm b}^{25}$ -45.6° (c 0.99, CHCl₂)

(3Z, 2R)-N-(tert-Butyloxy)carbonyl-2-amino-3-pentenethiol (6).

TFA (4.49 mL, 58.3 mmol) was added to a solution of **5** (1.00 g, 3.89 mmol) in water-saturated CH₂Cl₂ (250 mL). The reaction mixture was stirred for 2 h and quenched by the addition of H₂O (50 mL). The organic phase was washed repeatedly with H₂O (50 mL), dried and concentrated. Chromatography of the residue on silica gel (70 g, 15/1 hexane/EtOAc) gave **6** (543.0 mg, 64% yield) as a colorless oil: $[\alpha]_b^{25}$ +16.8° (c 0.485, CHCl₃); IR (CHCl₃) 3450, 2970, 2930, 1710, 1490, 1390, 1370, 1330, 1310, 1230, 1160, 1050, 1020, 860 cm⁻¹; H-NMR (500 MHz, CDCl₃) δ 1.45 (s 9H, (CH₃)₃C), 1.73 (dd, 3H, J=7.0, 1.8 Hz, CH₃-4), 2.65(ddd, 1H, J=13.4, 8.5, 6.5 Hz, H-1), 2.73 (ddd, 1H, J=13.4, 8.5, 4.6 Hz, H-1), 4.55 (m, 1H, H-2), 5.30 (ddq, 1H, J=10.8, 8.9, 1.8 Hz, H-3), 5.68 (dqd, 1H, J=10.8, 7.0, 1.1 Hz, H-4); ¹³C-NMR (125 MHz, CDCl₃) δ 13.60, 28.34 (x3), 29.98, 49.16, 79.54, 128.44, 128.71, 155.05; FABMS m/z: 218 (M+H)⁺; HRFABMS calcd. for C₁₀H₂₀O₂NS (M+H)⁺ 218.1215, found 218.1238; Anal. Calcd for C₁₀H₁₉O₂NS: C, 55.27; H, 8.81; N, 6.44; S, 14.75. Found: C, 55.42; H, 8.92; N, 6.33; S, 14.50.

(4S)-enantiomer was prepared according to the method above: $[\alpha]_0^{25}$ -16.0° (c 0.825, CHCl₃)

(4R)-2-[(1S, 2R)-2-methylcyclopropyl]-4-[(1Z)-1-propen-1-yl]thiazoline (+)-2a and (4R)-2-[(1R, 2S)-2-methylcyclopropyl]-4-[(1Z)-1-propen-1-yl]thiazoline (+)-2b.

Bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BOPCl, 2.32 g, 9.11 mmol) was added to a solution of 6 (1.59 g, 7.32 mmol), (±)-cis-2-methylcyclopropanecarboxylic acid 7 (795.4 mg, 7.94 mmol) and Et₂N (2.53 mL, 18.2 mmol) in dry CH₂Cl₂ (32 mL). The reaction mixture was stirred for 5 h and quenched by the addition of aqueous HCl (5%, 50 mL). The organic phase was washed with saturated aqueous NaHCO₃ and NaCl (50 mL), dried and concentrated. Chromatography of the residue on silica gel (165 g, 15/1 hexane/EtOAc) gave 8 (1.57 g, 72% yield) as a white gummy solid. A solution of the foregoing thiol ester 8 (1.54 g, 5.14 mmol) in CH₂Cl₂ (39 mL) was cooled to 0 °C and TFA (7.95 mL, 0.103 mol) was added to the solution. The reaction mixture was stirred for 1.5 h at 0 °C and concentrated under reduced pressure. Dry benzene (77 mL) was added to the resulting residue and concentrated twice for replacing the reaction solvent with benzene. A solution of the residue containing the corresponding thiol ester ammonium salt 9 in dry benzene (77 mL) was heated to reflux for 1 h and quenched by the addition of saturated aqueous NaHCO₃ (39 mL). The organic phase was washed with saturated aqueous NaCl (39 mL), dried and concentrated. Chromatography of the residue on silica gel (86 g, 20/1 to 5/1 hexane/EtOAc) gave a mixture of (+)-2a and (+)-2b (546.3 mg, 42% yield from 6). HPLC purification of the mixture (444.0 mg) using a TSK-gel, ODS-120T column (21.5 \(\phi \) x 600 mm, eluent: 35/65 MeCN/H₂O, detector: UV 235 nm, flow rate: 20 mL/min) gave (+)-2a (32.0 mg) and (+)-2b (45.2 mg) as a colorless oil, respectively: (+)-2a: $[\alpha]_{D}^{25}$ +141.6° (c 0.94, CHCl₃); IR (CHCl₃) 2970, 2900, 1620, 1440, 1390, 1290, 1180, 1140, 1120, 1080, 1010, 970, 920 cm⁻¹; H-NMR (500 MHz, C_6D_6) δ 0.72 (td, 1H, J=8.4, 4.3 Hz, H-3'), 0.96 (tq, 1H, J=8.4, 6.3 Hz, H-2'), 1.19 (d, 3H, J=6.3 Hz, CH₂-2'), 1.15-1.19 (m, 1H, H-3'), 1.45 (dd, 3H, J=6.9, 1.7 Hz, CH₃-2"), 1.64 (tdd, 1H, J=8.4, 5.5, 1.2 Hz, H-1'), 2.72 (dd, 1H,

J=10.8, 6.7 Hz, H-5), 3.07 (dd, 1H, J=10.8, 8.4 Hz, H-5), 5.21 (dddt, 1H, J=8.8, 8.4, 6.7, 1.2 Hz, H-4, observed as brq signal), 5.36 (dqd, 1H, J=10.7, 6.9, 1.2 Hz, H-2"), 5.51 (ddq, 1H, J=10.7, 8.8, 1.8 Hz, H-1"); 13 C-NMR (125 MHz, C₆D₆) δ 12.35 (CH₃-2'), 13.27 (CH₃-2"), 14.19 (C-3'), 16.02 (C-2'), 19.96 (C-1'), 39.90 (C-5), 73.88 (C-4), 125.53 (C-2"), 131.58 (C-1"), 168.06 (C-2); FABMS m/z: 182 (M+H)⁺; HRFABMS calcd. for C₁₀H₁₆NS (M+H)⁺ 182.1003, found 182.0992. (+)-**2b**: [α]_D²⁵ +108.4° (c 1.02, CHCl₃); IR (CHCl₃) 2970, 2900, 1620, 1440, 1390, 1290, 1180, 1150, 1130, 1080, 1020, 970, 920 cm⁻¹; ¹H-NMR (500 MHz, C₆D₆) δ 0.71 (td, 1H, J=8.4, 4.3 Hz, H-3'), 0.95 (tq, 1H, J=8.4, 6.3 Hz, H-2'), 1.17 (d, 3H, J=6.3 Hz, CH₃-2'), 1.14-1.18 (m, 1H, H-3'), 1.42 (dd, 3H, J=6.9, 1.8 Hz, CH₃-2"), 1.67 (td, 1H, J=8.4, 5.3 Hz, H-1'), 2.72 (dd, 1H, J=10.8, 9.8 Hz, H-5), 3.02 (dd, 1H, J=10.8, 8.3 Hz, H-5), 5.05 (dddd, 1H, J=9.8, 8.8, 8.3, 1.2 Hz, H-4, observed as brq signal), 5.40 (dqd, 1H, J=10.7, 6.9, 1.2 Hz, H-2"), 5.64 (ddq, 1H, J=10.7, 8.8, 1.8 Hz, H-1"); ¹³C-NMR (125 MHz, C₆D₆) δ 12.34 (CH₃-2'), 13.24 (CH₃-2"), 14.22 (C-3'), 15.97 (C-2'), 20.15 (C-1'), 39.77 (C-5), 74.00 (C-4), 125.61 (C-2"), 131.92 (C-1"), 168.34 (C-2); FABMS m/z: 182 (M+H)⁺; HRFABMS calcd. for C₁₀H₁₆NS (M+H)⁺ 182.1003, found 182.0995.

(4S)-2-[(1R, 2S)-2-methylcyclopropyl]-4-[(1Z)-1-propen-1-yl]thiazoline (-)-2a and (4S)-2-[(1S, 2R)-2-methylcyclopropyl]-4-[(1Z)-1-propen-1-yl]thiazoline (-)-2b.

By the procedure described for the preparation of (+)-2a and (+)-2b, (-)-6 was converted to a mixture of diastereomers of (-)-2 and HPLC purification of the mixture (98.3 mg) under the same condition shown above gave (-)-2a (25.2 mg) and (-)-2b (24.6 mg) as a colorless oil, respectively: (-)-2a: $[\alpha]_D^{25}$ -139.9° (c 0.90, CHCl₃) (-)-2b: $[\alpha]_D^{25}$ -104.3° (c 1.04, CHCl₃)

(4R)-2- $\{(1S, 2R)$ -2-methylcyclopropyl]-4- $\{(1Z)$ -1-propen-1-yl]thiazoline (+)-2a.

By the procedure described for the preparation of 2, (+)-6 (244.6 mg, 1.13 mmol) was converted to (+)-2a (70.3 mg, 34% yield) as a major product, using (+)-(1S, 2R)-2-methylcyclopropanecarboxylic acid (+)-7 ($[\alpha]_D^{25}$ +24.9° (c 0.495, 95% EtOH)) prepared according to the method of Bergman¹⁰. HPLC purification of the resulting residue (70.3 mg) under the same condition shown above gave (+)-2a (34.0 mg) as a colorless oil: $[\alpha]_D^{25}$ +145.6° (c 1.025, CHCl₃)

(2E)-3, 7-dimethyl-6, 7-epoxy-2-octen-1-ol (10).

The title compound was prepared according to the following procedure from geraniol. 16 Geraniol (25.00 g, 0.162 mol) was dissolved in CH₂Cl₂ (375 mL), acetone (375 mL) and buffered water (pH 7.5 adjusted by 0.2 M KH₂PO₄ and 0.2M Na₂HPO₄, 500 mL) and the reaction mixture was cooled to 0 °C. A freshly prepared solution of OXONE® (containing potassium peroxomonosulfate, KHSO₅, 29.89 g, 48.6 mmol) in H₂O (150 mL) was added dropwise to the biphasic mixture during 1 h at 0 °C. The reaction mixture was stirred for 2 h at 0 °C. The apparent pH of the mixture was monitored and kept constant at pH 7.5-8.0 by the addition of aqueous KOH (3 N) by use of a pH stat during the entire reaction time. Then the mixture was quenched by the addition of aqueous Na₂S₂O₃·5H₂O (4.02 g/H₂O 50 mL). The aqueous phase was extracted with CH₂Cl₂ (2x100mL), and the combined organic phases were dried and concentrated. Chromatography of the residue on silica gel (625 g, 3/1 to 2/1 hexane/EtOAc), followed by Kugelrohr distillation (160-180 °C, 1-2 mmHg) gave 10 (4.28 g, 16% yield) as a pale yellow oil, with recovery of geraniol (17.03 g, 68%) and a mixture of 10 and the 2, 3-epoxide (2.71 g). The same procedure with the recovered geraniol was carried out again and chromatography of the residue containing 10 on silica gel (2 times, 595 g and 164 g) gave 10 (10.66 g, 39% total yield), with recovery of geraniol (9.20 g, 37%): IR (CHCl₁) 3630, 3470 (broad), 2980, 2950, 2900, 1670, 1450, 1390, 1330, 1230, 1120, 1050, 1000, 870 cm⁻¹; H-NMR (500 MHz, CDCl₃) δ 1.27 (s, 3H, CH_3-7), 1.31 (s, 3H, H-8), 1.66 (m, 2H, H-5), 1.70 (d, 3H, J=1.4 Hz, CH_3-3), 2.18 (m, 2H, H-4), 2.71 (t, 1H, J=6.2 Hz, H-6), 4.17 (d, 2H, J=6.9 Hz, H-1), 5.46 (tq, 1H, J=6.9, 1.4 Hz, H-2); ¹³C-NMR (125 MHz, CDCl₃) δ 16.19, 18.68, 24.78, 27.09, 36.19, 58.35, 59.16, 64.00, 124.02, 138.41; FABMS m/z: 153 (M-OH)+; HRFABMS calcd. for $C_{10}H_{17}O$ (M-OH)* 153.1279, found 153.1276; Anal. Calcd for $C_{10}H_{18}O_2$: C, 70.55; H, 10.66. Found: C, 70.68; H, 10.76.

(2E)-3-Methyl-5-(2, 2, 5, 5-tetramethyl-1, 3-dioxolan-4-yl)-2-penten-1-ol (11).

A solution of **10** (10.43 g, 61.3 mmol) and TsOH·H₂O (11.65 g, 61.2 mmol) in aqueous acetone (9/1 acetone/H₂O, 260 mL) was stirred for 2 h. The reaction mixture was neutralized with aqueous NaHCO₃ (5.15 g, 61.3 mmol/H₂O 100mL) and concentrated. After the addition of NaCl (10 g), the resulting mixture was extracted with EtOAc (3x100 mL) and the combined organic phases were dried and concentrated. Chromatography of the residue on silica gel (2 times, 521 g and 319 g, 3/1 hexane/EtOAc), followed by Kugelrohr distillation (160-180 °C, 1-2 mmHg) gave **11** (9.43 g, 67% yield) as a colorless oil: IR (CHCl₃) 3630, 3000, 2950, 2900, 1450, 1380, 1370, 1220, 1200, 1120, 1060, 1000, 910, 850 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.08, 1.23, 1.31 and 1.40 (s, 12H, (CH₃)₂-2' and (CH₃)₂-5'), 1.48 (dddd, 1H, J=13.6, 10.8, 6.0, 3.4 Hz, H-5), 1.58-1.68 (m, 1H, H-5), 1.68 (d, 3H, J=1.2 Hz, CH₃-3), 2.05 (ddd, 1H, J=14.2, 10.4, 6.0 Hz, H-4), 2.25 (ddd, 1H, J=14.2, 10.8, 4.9 Hz, H-4), 3.64 (dd, 1H, J=9.5, 3.4 Hz, H-4'), 4.14 (d, 2H, J=6.9 Hz, H-1), 5.44 (tq, 1H, J=6.9, 1.2 Hz, H-2); ¹³C-NMR (125 MHz, CDCl₃) δ 16.32, 22.90, 26.01, 26.85, 27.53, 28.53, 36.62, 59.28, 80.10, 82.88, 106.55, 123.66, 139.00; FABMS m/z: 229 (M+H)⁺; HRFABMS calcd. for C₁₃H₂₅O₃ (M+H)⁺ 229.1804, found 229.1786; Anal. Calcd for C₁₃H₂₆O₃: C, 68.38; H, 10.60. Found: C, 68.24; H, 10.63.

(2E)-3-Methyl-5-(2, 2, 5, 5-tetramethyl-1, 3-dioxolan-4-yl)-2-penten-1-yl Triphenylphosphonium Bromide (12).

 CBr_4 (13.34 g, 40.2 mmol) and PPh_3 (10.55 g, 40.2 mmol) were added to a solution of 11 (7.99 g, 35.0 mmol) in dry CH₂Cl₂ (80 mL) at 0 °C, and the reaction mixture was stirred for 1 h at 0 °C. After evaporation of the solvent, the residue was triturated repeatedly with Et₂O (4x80 mL) and filtered. The combined filtrates were concentrated and another PPh₃ (13.76 g, 52.5 mmol) was added to a solution of the residue containing the corresponding bromide in dry benzene (80 mL). The reaction mixture was stirred for 16 h and filtered. The obtained precipitate was washed with Et₂O (2x250 mL) and dried under vacuum at 50 °C to give 12 (13.33 g, 69% yield from 11) as a white solid: mp 154.0-155.5 °C; IR (CHCl₃) 3670, 3400 (broad), 3050, 2960, 2870, 1620, 1590, 1440, 1370, 1220, 1200, 1120, 1000, 910, 850 cm⁻¹; H-NMR (500 MHz, CDCl₃) δ 1.04, 1.19, 1.26 (s, each 3H) and 1.38 (s, 6H, (CH_3) , -2', (CH_3) , -5' and CH_3 -3), 1.34-1.50 (m, 2H, H-5), 1.99 (ddd, 1H, J=19.5, 10.5, 4.5 Hz, H-4), 2.20 (ddd, 1H, J=19.5, 10.5, 4.5 Hz, H-4), 3.54 (dd, 1H, J=9.5, 3.0 Hz, H-4'), 4.69 (td, 1H, J=15.5, 7.5 Hz, H-1), 4.79 (td, 1H, J=15.5, 7.5 Hz, H-1), 5.19 (m, 1H, H-2), 7.68 (td, 6H, J=8.0, 3.5 Hz), 7.79 (tg, 3H, J=8.0, 1.5 Hz) and 7.89 (ddd, 6H, J=12.5, 8.0, 1.5 Hz, C₄H₄x3); ¹³C-NMR (125 MHz, CDCl₃) δ 17.00, 22.82, 24.40 (d, J=48.1 Hz), 25.95, 26.81, 27.41, 28.44, 37.13, 80.00, 82.81, 106.53, 108.65 (d, J=7.4 Hz), 118.25 (d, J=85.1 Hz, x3), 130.25 (d, J=11.1 Hz, x6), 133.82 (d, J=9.3 Hz, x6), 134.93 (x3), 146.63 (d, J=12.6 Hz); FABMS m/z: 473 (M-Br)⁺; HRFABMS calcd. for $C_{31}H_{32}O_{2}P$ (M-Br)⁺ 473.2609, found 473.2628; Anal. Calcd for C₃₁H₃₈O₂BrP·1/2H₂O: C, 66.19; H, 6.99. Found: C, 66.26; H, 6.81.

4-(4-Methoxybenzyl)oxy-1-butanol (14).

A solution of 1, 4-butanediol (7.00 g, 77.7 mmol) in anhydrous DMF (35 mL) was cooled to -15 °C and a solution of NaH (60% oil suspension, 3.42 g, 85.5 mmol) in anhydrous DMF (35 mL) was added. The reaction mixture was stirred for 0.5 h at -15 °C, and then a solution of 4-methoxybenzyl chloride (13.38 g, 85.4 mmol) in anhydrous DMF (21 mL) was added to the reaction mixture at the same temperature. After an additional 0.5 h, the reaction mixture was allowed to warm to 20 °C and stirred for another 1 h. It was quenched by slowly pouring it into ice-H₂O (195 mL) and extracted with Et₂O (3x130 mL). The combined organic phases were washed with saturated aqueous NaCl (3x130 mL), dried and concentrated. Chromatography of the residue on silica gel (350 g, 2/1 to 1/1 hexane/EtOAc), followed by Kugelrohr distillation (250 °C, 1-2 mmHg) gave 14 (13.66 g, 84% yield) as a colorless oil: IR (CHCl₃) 3620, 3440, 2950, 2870, 1610, 1590, 1510, 1460, 1360, 1300, 1230, 1170, 1090, 1030, 960, 820 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.64-1.74 (m, 4H, H-2 and H-3), 2.21 (t, 1H, J=6.0 Hz, OH), 3.50 (t, 2H, J=6.0 Hz, H-4), 3.64 (q, 2H, J=6.0 Hz, H-1), 3.80 (s, 3H, OCH₃), 4.45 (s, 2H, -CH₂O-), 6.88 and 7.26 (m, 4H, -C₆H₄-); ¹³C-NMR (125 MHz, CDCl₃) δ 26.69, 30.15, 55.21, 62.62, 70.00, 72.66, 113.78 (x2), 129.30 (x2), 130.18, 159.18; FABMS m/z: 210 (M)⁺; HRFABMS calcd. for C₁₂H₁₈O₃ (M)⁺ 210.1256, found 210.1288; Anal. Calcd for C₁₂H₁₈O₃: C, 68.55; H, 8.63. Found: C,

68.43; H, 8.59.

4-(4-Methoxybenzyl)oxy-1-butanal (15).

A mixture of dry CH₂Cl₂ (8 mL) and DMSO (890.6 mg, 11.4 mmol) was cooled below -50 °C and a solution of oxalyl chloride (964.6 mg, 7.60 mmol) in dry CH₂Cl₂ (4 mL) was added dropwise to the stirred cold solution. After 0.5 h at -50 °C, a solution of 14 (799.0 mg, 3.80 mmol) in dry CH₂Cl₂ (4 mL) was added dropwise to the reaction mixture at the same temperature. The mixture was stirred for 0.5 h at -50 °C, followed by the addition of Et₃N (2.12 mL, 15.2 mmol) dropwise. The reaction mixture was kept at or below -50 °C until the addition of Et₃N was complete, and it was allowed to warm slowly to 20 °C for 1.5 h and quenched by the addition of aqueous HCl (5%, 16 mL). The organic phase was washed with saturated aqueous NaHCO₃ and NaCl (16 mL), dried and concentrated. Kugelrohr distillation of the residue (200-210 °C, 1-2 mmHg) gave 15 (704.5 mg, 89% yield) as a pale yellow oil: IR (CHCl₃) 2950, 2850, 2750, 1720, 1610, 1590, 1510, 1460, 1360, 1300, 1230, 1170, 1090, 1030, 820 cm¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.93 (tt, 2H, J=7.0, 6.0 Hz, H-3), 2.53 (td, 2H, J=7.0, 1.8 Hz, H-2), 3.48 (t, 2H, J=6.0 Hz, H-4), 3.80 (s, 3H, OCH₃), 4.42 (s, 2H, -CH₂O-), 6.88 and 7.24 (m, 4H, -C₆H₄-), 9.77 (t, 1H, J=1.8 Hz, CHO); ¹³C-NMR (125 MHz, CDCl₃) δ 22.54, 40.93, 55.24, 68.81, 72.57, 113.77 (x2), 129.19 (x2), 130.33, 159.18, 202.24; FABMS m/z: 208 (M)⁺; HRFABMS calcd. for C₁₂H₁₆O₃ (M)⁺ 208.1099, found 208.1139; Anal. Calcd for C₁₂H₁₆O₃: C, 69.21; H, 7.74. Found: C, 69.08; H, 7.63.

(4E, 6E)-1-(4-Methoxybenzyl)oxy-7-methyl-9-(2, 2, 5, 5-tetramethyl-1, 3-dioxolan-4-yl)nona-4, 6-diene (13).

(a) Wittig reaction of 12 with 15: A suspension of 12 (12.93 g, 23.4 mmol) in dry THF (390 mL) was cooled to -78 °C and n-BuLi (14.60 mL, 1.6 M in hexane, 23.4 mmol) was added dropwise to the suspension. The reaction mixture was stirred for 0.5 h at -78 °C. A solution of 15 (6.32 g, 30.3 mmol) in dry THF (64 mL) was added dropwise to the resulting dark red solution at -78 °C over 20 min. The reaction mixture was allowed to warm slowly to 20 °C for 16 h and quenched slowly by the addition of saturated aqueous NH₄Cl (650 mL). The resulting mixture was extracted with EtOAc (800 and 2x600 mL) and the combined organic phases were washed with saturated aqueous NaCl (650 mL), dried and concentrated to afford a white insoluble solid. The solid was washed with hexane/EtOAc (20/1, 100 mL) and the washing solution was concentrated. Chromatography of the residue on silica gel (650 g, 15/1 to 10/1 hexane/EtOAc) gave 13 (6.87 g, 73% yield, E/Z=1.9/1) as a pale yellow oil. HPLC purification of the mixture (6.87 g) using Senshu Pak. SSC-Silica-5251-N and Shiseido Silica SG120Å column (each 20 \(\phi \) x 250 mm, eluent: CH₂Cl₂, detector: UV 254 nm, flow rate: 30-35 mL/min) gave 13 (3.46 g, E/Z=99.8/0.2) and the corresponding (4Z)-isomer (2.16 g, E/Z=6/94) as a pale yellow oil, respectively: 13: IR (CHCl₂) 2950, 2860, 1610, 1590, 1520, 1450, 1370, 1220, 1170, 1110, 1040, 1000, 960, 910, 820 cm⁻¹; H-NMR (500 MHz, CDCl₁) δ 1.09, 1.24, 1.33, 1.42 and 1.75 (s, 15H, $(CH_3)_{7}$ -2', $(CH_3)_{7}$ -5' and CH_3 -7), 1.50 (dddd, 1H, J=13.8, 10.5, 6.5, 3.5 Hz, H-9), 1.60-1.68 (m, 1H, H-9), 1.70 (tt, 2H, J=7.0, 6.5 Hz, H-2, observed as quint signal), 2.07 (ddd, 1H, J=14.0, 10.5, 6.5 Hz, H-8), 2.18 (qd, 2H, J=7.0, 1.5 Hz, H-3, observed as brq signal), 2.27 (ddd, 1H, J=14.0, 10.5, 5.0 Hz, H-8), 3.46 (t, 2H, J=6.5 Hz, H-1), 3.66 (dd, 1H, J=9.5, 3.5 Hz, H-4'), 3.80 (s, 3H, OCH₃), 4.43 (s, 2H, -CH₂O-), 5.58 (dt, 1H, J=15.0, 7.0 Hz, H-4), 5.83 (d, 1H, J=11.0 Hz, H-6), 6.25 (ddt, 1H, J=15.0, 11.0, 1.5 Hz, H-5), 6.88 and 7.26 (m, 4H, $-C_6H_4$ -); ^{13}C -NMR (125 MHz, CDCl₃) δ 16.56, 22.91, 26.03, 26.82, 27.72, 28.54, 29.48, 29.57, 36.84, 55.22, 69.47, 72.52, 80.07, 82.87, 106.47, 113.72 (x2), 124.94, 126.93, 129.18 (x2), 130.69, 131.88, 135.66, 159.09; FABMS m/z: 402 (M)*; HRFABMS calcd. for C₂₅H₃₈O₄ (M)* 402.2770, found 402.2758; Anal. Calcd for C₂₅H₂₈O₄: C, 74.59; H, 9.51. Found: C, 74.53; H, 9.51.

(4Z)-isomer: IR (CHCl₃) 2950, 2860, 1610, 1580, 1520, 1450, 1370, 1220, 1170, 1110, 1030, 1000, 910, 820 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.08, 1.22, 1.31, 1.41 and 1.75 (s, 15H, (CH₃)₂-2', (CH₃)₂-5' and CH₃-7), 1.49 (dddd, 1H, J=13.8, 10.5, 6.5, 3.5 Hz, H-9), 1.60-1.68 (m, 1H, H-9), 1.68 (tt, 2H, J=7.5, 7.0 Hz, H-2, observed as quint signal), 2.11 (ddd, 1H, J=14.0, 10.0, 6.5 Hz, H-8), 2.24 (q, 2H, J=7.5 Hz, H-3), 2.29 (ddd, 1H, J=14.0, 10.5, 4.5 Hz, H-8), 3.44 (t, 2H, J=7.0 Hz, H-1), 3.64 (dd, 1H, J=9.5, 3.5 Hz, H-4'), 3.78 (s,

3H, OCH₃), 4.41 (s, 2H, -CH₂O-), 5.34 (dt, 1H, J=10.5, 7.5 Hz, H-4), 6.12 (d, 1H, J=11.5 Hz, H-6), 6.18 (dd, 1H, J=11.5, 10.5, H-5, observed as t signal), 6.85 and 7.24 (m, 4H, -C₆H₄-); ¹³C-NMR (125 MHz, CDCl₃) δ 16.35, 22.91, 24.19, 26.01, 26.84, 27.78, 28.54, 29.72, 37.35, 55.24, 69.47, 72.54, 80.06, 82.79, 106.49. 113.74 (x2), 120.37, 125.00, 129.16 (x2), 129.47, 130.71, 137.83, 159.09; FABMS m/z: 402 (M)⁺; HRFABMS calcd. for C₂₅H₃₈O₄ (M)⁺ 402.2770, found 402.2728; Anal. Calcd for C₂₅H₃₈O₄: C, 74.59; H, 9.51. Found: C, 74.90; H, 9.52.

(b) Wittig-Horner reaction of 16 with 15: A solution of t-BuOK (161.0 mg, 1.43 mmol) in dry THF (2 mL) was added to a mixed solution of 15 (300.0 mg, 1.44 mmol) and 16 (200.0 mg, 0.574 mmol) in dry THF (2 mL). The reaction mixture was stirred vigorously for 1.5 h and quenched by the addition of saturated aqueous NaCl (8 mL). The aqueous phase was extracted with EtOAc (3x8 mL), and the combined organic phases were washed with saturated aqueous NaCl (8 mL), dried and concentrated. Chromatography of the residue on silica gel (10 g, 20/1 to 1/2 hexane/EtOAc) gave 13 (117.9 mg, 51% yield, E/Z=8.5/1) as a pale yellow oil, with recovery of 16 (26.0 mg, 13%).

Diethyl (2E)-[3-methyl-5-(2, 2, 5, 5-tetramethyl-1, 3-dioxolan-4-yl)-2-penten-1-yl] phosphonate (16).

CBr₄ (4.40 g, 13.3 mmol) and PPh₃ (3.48 g, 13.3 mmol) were added to a solution of 11 (2.52 g, 11.0 mmol) in dry CH₂Cl₂ (25 mL) at 0 °C, and the reaction mixture was stirred for 1 h at 0 °C. After evaporation of the solvent, the residue was triturated repeatedly with Et₂O (4x25 mL) and filtered. The combined filtrates were concentrated and triethyl phosphite ((EtO)₃P, 5.68 mL, 33.1 mmol) was added to a solution of the residue containing the corresponding bromide in dry benzene (10 mL). The reaction mixture was heated to reflux for 2.5 h and distilled at 160-180 °C (20 mmHg) to remove low-boiling substances. Chromatography of the residue on silica gel (125 g, 1/2 hexane/EtOAc), followed by Kugelrohr distillation (250 °C, 1-2 mmHg) twice gave 16 (2.88 g, 75% yield from 11) as a colorless oil: IR (CHCl₂) 3400 (broad), 3000, 2950, 2870, 1450, 1370, 1240, 1110, 1030, 1000, 970, 850 cm $^{-1}$; H-NMR (500 MHz, CDCl₃) δ 1.07, 1.22, 1.30 and 1.39 (s, 12H, (CH₂)₀-2' and (CH₂)₀-5'), 1.29 (t, 6H, J=7.5 Hz, (OCH₂CH₃)₀), 1.47 (dddd, 1H, J=13.5, 10.5, 6.5, 3.5 Hz, H-5), 1.57-1.66 (m, 1H, H-5), 1.65 (d, 3H, J=3.5 Hz, CH₃-3), 2.07 (m, 1H, H-4), 2.25 (ddd, 1H, J=19.3, 10.5, 5.0 Hz, H-4), 2.55 (dd, 2H, J=21.5, 7.5 Hz, H-1), 3.64 (dd, 1H, J=9.5, 3.5 Hz, H-4'), 4.03-4.11 (m, 4H, (OCH,CH,)₂), 5.22 (qq, 1H, J=7.5, 1.5 Hz, H-2); ¹³C-NMR (125 MHz, CDCl₁) δ 16.31, 16.41, 16.45, 22.87, 26.00, 26.35 (d, J=140.5 Hz), 26.81, 27.53, 28.51, 36.66, 61.68 (x2), 80.02, 82.65, 106.47, 112.98 (d, J=9.3 Hz), 139.47 (d, J=14.7 Hz); FABMS m/z: 349 (M+H)⁺; HRFABMS calcd. for $C_{17}H_{14}O_5P$ (M+H)⁺ 349.2144, found 349.2136; Anal. Calcd for C₁₇H₃₃O₅P: C, 58.60; H, 9.54; P, 8.89. Found: C, 58.35; H, 9.45; P, 8.86.

(4E, 6E)-10, 11-Dihydroxy-7, 11-dimethyl-1-(4-methoxybenzyl)oxydodeca-4, 6-diene (17).

A solution of 13 (3.31 g, 8.22 mmol) and TsOH·H₂O (938.4 mg, 4.93 mmol) in aqueous MeOH (9/1 MeOH/H₂O, 83 mL) was stirred for 5 h. The reaction mixture was neutralized with aqueous NaHCO₃ (415.0 mg, 4.94 mmol/H₂O 20 mL) and concentrated. After the addition of saturated aqueous NaCl (30 mL), the resulting mixture was extracted with EtOAc (3x50 mL), and the combined organic phases were dried and concentrated. Chromatography of the residue on silica gel (165 g, 2/1 hexane/EtOAc) gave 17 (2.75 g, 92% yield) as a pale yellow oil, with recovery of 13 (318.7 mg, 10%). The same procedure with the recovered 13 was carried out again to afford 17 (2.95 g) in 99% total yield from 13: IR (CHCl₃) 3580, 2950, 2870, 1620, 1590, 1510, 1440, 1360, 1300, 1240, 1170, 1090, 1040, 970, 820 cm⁻¹; H-NMR (500 MHz, CDCl₃) δ 1.16, 1.21 and 1.74 (s, 9H, CH₃-7, CH₃-11 and H-12), 1.44 (dddd, 1H, J=14.0, 10.5, 9.0, 5.5 Hz, H-9), 1.62 (dddd, 1H, J=14.0, 9.5, 7.0, 1.5 Hz, H-9), 1.70 (tt, 2H, J=7.0, 6.5 Hz, H-2, observed as quint signal), 2.08-2.17 (m, 1H, H-8), 2.18 (qd, 2H, J=7.0, 1.5 Hz, H-3, observed as brd signal), 2.31 (ddd, 1H, J=14.0, 9.5, 5.5 Hz, H-8), 3.35 (dd, 1H, J=10.5, 1.5 Hz, H-10, observed as brd signal), 3.46 (t, 2H, J=6.5 Hz, H-1), 3.80 (s, 3H, OCH₃), 4.43 (s, 2H, -CH₂O-), 5.58 (dt, 1H, J=15.0, 7.0 Hz, H-4), 5.85 (d, 1H, J=11.0 Hz, H-6), 6.24 (ddt, 1H, J=15.0, 1.0, 1.5 Hz, H-5), 6.88 and 7.26 (m, 4H, -C₆H₄-); ¹³C-NMR (125 MHz, CDCl₃) δ 16.44, 23.24, 26.45, 29.49, 29.56, 29.69, 36.86, 55.25, 69.47, 72.51, 73.04, 78.17, 113.74 (x2), 125.19, 126.86, 129.20 (x2), 130.71,

132.03, 136.06, 159.09; FABMS m/z: 362 (M)⁺; HRFABMS calcd. for $C_{22}H_{34}O_4$ (M)⁺ 362.2457, found 362.2410; Anal. Calcd for $C_{22}H_{34}O_4$: C, 72.89; H, 9.45. Found: C, 72.67; H, 9.62.

(4E, 6E)-9-Formyl-1-(4-methoxybenzyl)oxy-7-methylnona-4, 6-diene (18).

NaIO₄ (3.27 g, 15.3 mmol) was added to a solution of 17 (2.22 g, 6.12 mmol) in a mixture of acetone (111 mL) and H₂O (22 mL). The reaction mixture was stirred vigorously for 2 h and quenched by the addition of saturated aqueous NaCl (220 mL). The resulting mixture was extracted with EtOAc (3x220 mL) and the combined organic phases were washed with saturated aqueous NaCl (110 mL), dried and concentrated. Chromatography of the residue on silica gel (220 g, 4/1 to 1/1hexane/EtOAc) gave 18 (1.43 g, 77% yield) as a pale yellow oil, with recovery of 17 (598.5 mg, 27%). The same procedure with the recovered 17 was repeated two more times to afford 18 (1.74 g) in 94% total yield from 17: IR (CHCl₃) 2950, 2850, 2730, 1720, 1610, 1590, 1510, 1440, 1360, 1240, 1170, 1100, 1040, 970, 900, 820 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.70 (tt, 2H, J=7.0, 6.5 Hz, H-2, observed as quint signal), 1.74 (s, 3H, CH₃-7), 2.18 (dtd, 2H, J=7.5, 7.0, 1.5 Hz, H-3, observed as brq signal), 2.37 (t, 2H, J=7.5 Hz, H-8), 2.55 (td, 2H, J=7.5, 1.5 Hz, H-9), 3.45 (t, 2H, J=6.5 Hz, H-1), 3.80 (s, 3H, OCH₂), 4.43 (s, 2H, -CH₂O-), 5.59 (dt, 1H, J=15.0, 7.5 Hz, H-4), 5.80 (d, 1H, J=10.5 Hz, H-6), 6.22 (ddt, 1H, J=15.0, 10.5, 1.5 Hz, H-5), 6.88 and 7.26 (m, 4H, -C_xH_x-), 9.77 (t, 1H, J=1.5 Hz, CHO); ¹³C-NMR (125 MHz, CDCl₃) δ 16.53, 29.47 (x2), 31.79, 41.98, 55.19, 69.40, 72.49, 113.69 (x2), 125.44, 126.58, 129.15 (x2), 130.66, 132.68, 133.87, 159.06, 202.15; FABMS m/z: 302 (M)+; HRFABMS calcd. for $C_{19}H_{26}O_3$ (M)⁺ 302.1882, found 302.1860; Anal. Calcd for $C_{19}H_{26}O_3$: C, 75.46; H, 8.67. Found: C, 75.50; H, 8.66.

(4E, 6E, 10R)-10-hydroxy-1-(4-methoxybenzyl)oxy-7-methyltrideca-4, 6, 12-triene (20).

The following procedure is a modification of that reported by Hafner et al. 13 Allylmagnesium chloride (8.02 mL, 2.0 M in THF, 16.0 mmol) was added dropwise over 20 min at 0 °C to a 82 mM solution of [(4R, 5R)-2, 2-dimethyl-1, 3-dioxolane-4, 5-bis(diphenylmethoxy)]cyclopentadienyl-chlorotitanium, (R, R)-19 (10.92 g, 17.8 mmol) in dry THF (217 mL). The reaction mixture was stirred for 1 h at 0 °C and the slightly orange suspension was formed. A solution of 18 (1.80 g, 5.95 mmol) in dry THF (15 mL) was added to the suspension at -78 °C over 20 min. The reaction mixture was stirred for 1.5 h at -78 °C and quenched by the addition of aqueous NH₄F (45%, 36 mL). After hydrolysis with aqueous NH₄F for 12 h at 20 °C, the resulting mixture was filtered through Celite and extracted with EtOAc (2x200 mL). The combined organic phases were washed with saturated aqueous NaCl (2x100 mL), dried and concentrated. Chromatography of the residue on silica gel (2 times, 2x360 g, 8/1 to 5/1 hexane/EtOAc and CH₂Cl, to 20/1 CH₂Cl,/EtOAc) gave 20 (1.95 g, 95% yield) as a colorless oil: $[\alpha]_{25}^{15} + 3.6^{\circ}$ (c 0.985, CHCl₃); IR (CHCl₃) 3580, 2950, 2850, 1610, 1510, 1440, 1360, 1300, 1240, 1170, 1090, 1030, 960, 910 cm⁻¹; H-NMR (500 MHz, CDCl₂) δ 1.53-1.65 (m, 2H, H-9), 1.70 (tt, 2H, J=7.5, 6.5 Hz, H-2, observed as quint signal), 1.73 (d, 3H, J=0.5 Hz, CH₃-7), 2.06-2.24 (m, 3H, H-8 and H-11), 2.17 (qd, 2H, J=7.5, 1.5 Hz, H-3, observed as brq signal), 2.30 (dddt, 1H, J=13.8, 6.5, 4.5, 1.5 Hz, H-11), 3.45 (t, 2H, J=6.5 Hz, H-1), 3.63 (ddd, 1H, J=12.0, 8.0, 4.5 Hz, H-10), 3.80 (s, 3H, OCH₃), 4.43 (s, 2H, -CH₂O-), 5.12 and 5.14 (m, 2H, H-13), 5.57 (dt, 1H, J=15.0, 7.5 Hz, H-4), 5.78-5.87 (m, 1H, H-12), 5.83 (d, 1H, J=10.5 Hz, H-6), 6.24 (ddt, 1H, J=15.0, 10.5, 1.5 Hz, H-5), 6.87 and 7.26 (m, 4H, -C_eH_e-); ¹³C-NMR (125 MHz, CDCl₃) δ 16.47, 29.47, 29.56, 34.79, 35.93, 41.93, 55.22, 69.46, 70.37, 72.49, 113.71 (x2), 118.08, 124.91, 126.90, 129.18 (x2), 130.68, 131.84, 134.71, 136.12, 159.06; FABMS m/z: 344 (M)⁺, 343 (M-H)⁺; HRFABMS calcd. for $C_{22}H_{31}O_3$ (M-H)⁺ 343.2273, found 343.2264; Anal. Calcd for $C_{22}H_{32}O_3$: C, 76.70; H, 9.36. Found: C, 76.20; H, 9.22.

[(4E, 6E, 10R)-1-(4-methoxybenzyl)oxy-7-methyltrideca-4, 6, 12-trien-10-yl] (R)- α -methoxy- α -(trifluoromethyl)phenyl acetate (21), the Mosher ester of 20.

Pyridine (120 μ L, 1.48 mmol) and (S)-(+)- α -methoxy- α -(trifluoromethyl)phenylacetyl chloride (29 μ L, 0.155 mmol) were added to a solution of **20** (10.5 mg, 0.030 mmol) in dry CH₂Cl₂ (0.2 mL). The reaction mixture was stirred for 0.5 h, quenched by the addition of aqueous HCl (5%, 1 mL) and extracted with EtOAc

(2x3 mL). The combined organic phases were washed with saturated aqueous NaHCO₃ and NaCl (2 mL), dried and concentrated. Preparative TLC purification of the residue (3/1 hexane/EtOAc) gave **21** (11.3 mg, 66% yield) as a colorless oil: 1 H-NMR (500 MHz, CDCl₃) δ 1.64 (d, 3H, J=1.0 Hz, CH₃-7), 1.65-1.74 (m, 4H, H-2 and H-9), 1.92 (m, 2H, H-8), 2.17 (qd, 2H, J=7.0, 1.5 Hz, H-3, observed as brq signal), 2.43 (t, 2H, J=7.0 Hz, H-11), 3.45 (t, 2H, J=6.5 Hz, H-1), 3.56 (m, 3H, OCH₃ of MTPA), 3.80 (s, 3H, OCH₃ of PMB), 4.43 (s, 2H, -CH₂O-), 5.08-5.16 (m, 3H, H-10 and H-13), 5.55 (dt, 1H, J=15.0, 7.0 Hz, H-4), 5.67 (dq, 1H, J=11.0, 1.0 Hz, H-6), 5.75 (ddt, 1H, J=16.5, 10.5, 7.0 Hz, H-12), 6.20 (ddt, 1H, J=15.0, 11.0, 1.5 Hz, H-5), 6.87 and 7.26 (m, 4H, -C₆H₄- of PMB), 7.39 and 7.55 (m, 5H, C₆H₅ of MTPA); 13 C-NMR (125 MHz, CDCl₃) δ 16.35, 29.50, 29.56, 31.48, 34.88, 38.19, 55.25, 55.53, 69.49, 72.53, 76.18, 84.45 (q), 113.74 (x2), 118.53, 123.37 (q, J=288.2 Hz), 125.16, 126.78, 127.34 (x2), 128.31 (x2), 129.21 (x2), 129.53, 130.71, 132.16, 132.37, 132.96, 134.90, 159.09, 166.17; FABMS m/z; 560 (M)⁺.

(4E, 6E, 10R)-10-Methoxy-1-(4-methoxybenzyl)oxy-7-methyltrideca-4, 6, 12-triene (22).

A solution of 20 (1.90 g, 5.52 mmol) in anhydrous DMF (19 mL) was cooled to 0 °C and NaH (60% oil suspension, 441.4 mg, 11.0 mmol) was added to the solution. The reaction mixture was stirred for 0.5 h at 0 °C, and then a solution of MeI (1.03 mL, 16.5 mmol) in anhydrous DMF (1.9 mL) was added to the mixture at the same temperature. The reaction mixture was allowed to warm to 20 °C and stirred for 2.5 h. It was quenched by slowly pouring it into cold saturated aqueous NaCl (380 mL) and extracted with EtOAc (3x190 mL). The combined organic phases were washed with saturated aqueous NaCl (190 mL), dried and concentrated. Chromatography of the residue on silica gel (190 g, 20/1 hexane/EtOAc) gave 22 (1.76 g, 89% yield) as a colorless oil: $[\alpha]_{5}^{p_{5}}$ -2.1° (c 1.015, CHCl₃); IR (CHCl₃) 2950, 2850, 1640, 1610, 1510, 1450, 1360, 1300, 1240, 1170, 1090, 1030, 960, 890 cm⁻¹; ¹H-NMR (500 MHz, CDCl₂) δ 1.59 (td, 2H, J=8.0, 6.0 Hz, H-9), 1.70 (tt, 2H, J=7.0, 6.5 Hz, H-2, observed as quint signal), 1.72 (s, 3H, CH₂-7), 2.01-2.16 (m, 2H, H-8), 2.17 (qd, 2H, J=7.0, 1.5 Hz, H-3, observed as brq signal), 2.27 (m, 2H, H-11), 3.19 (quint, 1H, J=6.0 Hz, H-10), 3.34 (s, 3H, OCH₂-10), 3.45 (t, 2H, J=6.5 Hz, H-1), 3.80 (s, 3H, OCH₂ of PMB), 4.43 (s, 2H, -CH₂O-), 5.04-5.11 (m, 2H, H-13), 5.56 (dt, 1H, J=15.0, 7.0 Hz, H-4), 5.78-5.82 (m, 1H, H-6), 5.81 (ddt, 1H, J=17.0, 10.0, 7.0 Hz, H-12), 6.24 (ddt, 1H, J=15.0, 10.5, 1.5 Hz, H-5), 6.87 and 7.26 (m, 4H, $-C_6H_4$ -); ^{13}C -NMR (125 MHz, CDCl₃) δ 16.53, 29.50, 29.60, 31.59, 35.34, 37.65, 55.25, 56.53, 69.52, 72.53, 79.90, 113.74 (x2), 116.93, 124.72, 127.03, 129.19 (x2), 130.72, 131.61, 134.75, 136.34, 159.09; FABMS m/z: 358 (M)⁺, 357 (M-H)⁺; HRFABMS calcd. for C₂₁H₃₁O₃ (M-H)⁺ 357.2430, found 357.2382; Anal. Calcd for C₂₁H₂₄O₃: C, 77.05; H, 9.56. Found: C, 77.21; H, 9.55.

(4E, 6E, 10R)-10-Methoxy-7-methyltrideca-4, 6, 12-trien-1-ol (23).

Me₂S (3.24 mL, 44.1 mmol) and MgBr₂·OEt₂ (3.41 g, 13.2 mmol) were added to a solution of **22** (1.52 g, 4.41 mmol) in dry CH₂Cl₂ (152 mL). The reaction mixture was stirred vigorously for 2 h, quenched by the addition of saturated aqueous NH₄Cl (100 mL) and extracted with CH₂Cl₂ (3x50 mL). The combined organic phases were washed with saturated aqueous NaCl (100 mL), dried and concentrated. Chromatography of the residue on silica gel (150 g, 8/1 to 5/1 hexane/EtOAc) gave **23** (0.24 g, 23% yield) as a pale yellow oil, with recovery of **22** (1.09 g, 72%). The same procedure with the recovered **22** was repeated four more times to afford **23** (802.6 mg) in 76% total yield from **22**, with recovery of **22** (89.0 mg. 6%): $[\alpha]_{\rm D}^{15}$ -2.8° (*c* 2.53, CHCl₃), lit. $[\alpha]_{\rm D}^{15}$ -1.4° (*c* 2.90, CDCl₃)^{6a}, IR (CHCl₃) 3450 (broad), 2940, 1640, 1610, 1510, 1440, 1350, 1090, 960, 910 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.60 (td, 2H, J=8.0, 6.0 Hz, H-9), 1.68 (tt, 2H, J=7.0, 6.5 Hz, H-2, observed as quint signal), 1.73 (s, 3H, CH₃-7), 2.01-2.16 (m, 2H, H-8), 2.19 (qd, 2H, J=7.0, 1.5 Hz, H-3, observed as brq signal), 2.27 (m, 2H, H-11), 3.20 (quint, 1H, J=6.0 Hz, H-10), 3.34 (s, 3H, OCH₃-10), 3.67 (t, 2H, J=6.5 Hz, H-1), 5.04-5.11 (m, 2H, H-13), 5.58 (dt, 1H, J=15.0, 7.0 Hz, H-4), 5.80 (ddt, 1H, J=17.3, 10.0, 7.0 Hz, H-12), 5.81 (d, 1H, J=10.5 Hz, H-6), 6.28 (ddt, 1H, J=15.0, 10.5, 1.5 Hz, H-5); ¹³C-NMR (125 MHz, CDCl₃) δ 16.56, 29.22, 31.59, 32.47, 35.35, 37.66, 56.54, 62.54, 79.90, 116.96, 124.59, 127.25, 131.36, 134.75, 136.66; FABMS m/z: 238 (M)*; HRFABMS calcd. for C₁₅H₂₆O₂ (M)* 238.1933, found 238.1918.

(4E, 6E, 10R)-1-Iodo-10-methoxy-7-methyltrideca-4, 6, 12-triene (24).

Methanesulfonyl chloride (414 μL, 5.35 mmol) was added to a solution of 23 (637.4 mg, 2.67 mmol) in pyridine (6.4 mL) at 0 °C and the reaction mixture was stirred for 1 h at 0 °C. The resulting mixture was quenched by the addition of H₂O (30 mL) and extracted with EtOAc (30 mL). The organic phase was washed with aqueous HCl (5%, 2x30 mL), saturated aqueous NaHCO₃ and NaCl (30 mL), dried and concentrated. Chromatography of the residue on silica gel (64 g, 5/1 hexane/EtOAc) gave the corresponding mesylate (767.8 mg, quant.) as a colorless oil. NaI (1.62 g, 10.8 mmol) was added to a solution of the foregoing mesylate (767.8 mg, 2.70 mmol) in acetone (23 mL). The reaction mixture was heated to reflux for 2 h and concentrated. After the addition of saturated aqueous NaCl (40 mL), the resulting mixture was extracted with EtOAc (2x40 mL), dried and concentrated. Chromatography of the residue on silica gel (77 g, 40/1 hexane/EtOAc) gave 24 (809.5 mg, 87% yield from 23) as a pale brown oil: $[\alpha]_{12}^{15}$ -2.5° (c 4.05, CHCl₃), lit. $[\alpha]_{13}^{15}$ -0.9° (c 4.35, CHCl₂)^{6a}; IR (CHCl₄) 2930, 1640, 1430, 1360, 1160, 1090, 990, 960, 910 cm⁻¹; H-NMR (500 MHz, CDCl₂) δ 1.60 (td, 2H, J=8.0, 6.0 Hz, H-9), 1.74 (s, 3H, CH₃-7), 1.91 (tt, 2H, J=7.0, 6.5 Hz, H-2, observed as quint signal), 2.02-2.16 (m, 2H, H-8), 2.21 (qd, 2H, J=7.0, 1.5 Hz, H-3, observed as brq signal), 2.27 (m, 2H, H-11), 3.17-3.23 (m, 1H, H-10), 3.19 (t, 2H, J=6.5 Hz, H-1), 3.34 (s, 3H, OCH,-10), 5.04-5.12 (m, 2H, H-13), 5.49 (dt, 1H, J=15.0, 7.0 Hz, H-4), 5.80 (d, 1H, J=10.5 Hz, H-6), 5.81 (ddt, 1H, J=17.3, 10.0, 7.0 Hz, H-12), 6.30 (ddt, 1H, J=15.0, 10.5, 1.5 Hz, H-5); ¹³C-NMR (125 MHz, CDCl₂) δ 6.56, 16.59, 31.59, 33.07, 33.44, 35.37, 37.66, 56.59, 79.90, 116.96, 124.47, 128.13, 129.52, 134.74, 137.15; FABMS m/z: 348 (M)⁺; HRFABMS calcd. for $C_{15}H_{25}OI(M)^{+}$ 348.0950, found 348.0953.

(4E, 6E, 10R)-10-Methoxy-7-methyltrideca-4, 6, 12-trien-1-yl Triphenylphosphonium Iodide (25).

PPh₃ (2.42 g, 9.22 mmol) was added to a solution of **24** (803.4 mg, 2.31 mmol) in acetonitrile (24 mL). The reaction mixture was heated to reflux for 7 h and concentrated. The residue was triturated repeatedly with Et₂O (3x50 mL) and filtered. The precipitate was dried under vacuum to afford **25** (1.41 g, quant.) as a gummy white solid: $[\alpha]_D^{25}$ -0.5° (c 7.20, CHCl₃), lit. $[\alpha]_D^{23}$ -0.0° (c 7.0, CHCl₃)^{6a}; IR (CHCl₃) 2930, 1640, 1590, 1440, 1110, 995, 965, 915 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.57 (m, 2H, H-9), 1.71 (s, 3H, CH₃-7), 1.75 (m, 2H, H-2), 1.99-2.15 (m, 2H, H-8), 2.26 (m, 2H, H-11), 2.49 (m, 2H, H-3), 3.18 (quint, 1H, J=6.0 Hz, H-10), 3.33 (s, 3H, OCH₃-10), 3.74 (m, 2H, H-1), 5.03-5.10 (m, 2H, H-13), 5.42 (dt, 1H, J=15.0, 7.5 Hz, H-4), 5.75 (d, 1H, J=11.0 Hz, H-6), 5.79 (ddt, 1H, J=17.3, 10.0, 7.0 Hz, H-12), 6.31 (dd, 1H, J=15.0, 11.0 Hz, H-5), 7.71 and 7.82 (m, 15 H, C₆H₅x3); ¹³C-NMR (125 MHz, CDCl₃) δ 16.71, 22.18, 22.67, 31.55, 33.09 (d, J=16.6 Hz), 35.34, 37.59, 56.51, 79.84, 116.94, 117.81, 118.49, 124.23, 129.08 (d, J=16.7 Hz, x3), 130.50 (d, J=12.1 Hz, x6), 133.71 (d, J=9.2 Hz, x6), 134.67, 135.05 (x3), 137.93; FABMS m/z: 483 (M-I)⁺; HRFABMS calcd. for C₃₃H₄₀OP (M-I)⁺ 483.2817, found 483.2791.

(2Z, 6E, 8E, 12R)-12-Methoxy-9-methylpentadeca-2, 6, 8, 14-tetraene (26).

A solution of **25** (81.9 mg, 0.134 mmol) in dry THF (1.64 mL) was cooled to -78 °C and LiHMDS (240 μ L, 0.5 M in THF, 0.120 mmol) was added dropwise to the solution. The reaction mixture was stirred for 0.5 h at -78 °C. A solution of acetaldehyde (29.5 mg, 0.670 mmol) in dry THF (0.2 mL) was added dropwise to the resulting red solution at -78 °C over 20 min. The reaction mixture was allowed to warm slowly to 0 °C for 1.5 h and quenched by the addition of saturated aqueous NH₄Cl (5 mL). The resulting mixture was extracted with EtOAc (2x10 mL) and the combined organic phases were dried and concentrated. Chromatography of the residue on silica gel (17 g, 3/1 to 2/1 hexane/CH₂Cl₂), followed by preparative TLC purification (benzene) gave **26** (13.6 mg, 41% yield) as a colorless oil: $[\alpha]_p^{15}$ -2.9° (*c* 0.31, CHCl₃); IR (CHCl₃) 2940, 2860, 1640, 1430, 1360, 1260, 1090, 970, 910 cm⁻¹; H-NMR (500 MHz, CDCl₃) δ 1.60 (td, 2H, J=8.0, 6.0 Hz, H-11), 1.61 (dd, 3H, J=6.0, 1.5 Hz, H-1), 1.73 (s, 3H, CH₃-9), 2.02-2.17 (m, 6H, H-4, H-5 and H-10), 2.27 (m, 2H, H-13), 3.20 (q, 1H, J=6.0 Hz, H-12), 3.34 (s, 3H, OCH₃-12), 5.04-5.12 (m, 2H, H-15), 5.40 (m, 1H, H-2), 5.46 (dtq, 1H, J=11.0, 7.0, 1.5 Hz, H-3), 5.59 (dt, 1H, J=15.0, 6.5 Hz, H-6), 5.81 (ddt, 1H, J=17.3, 10.5, 7.0 Hz, H-14), 5.81 (d, 1H, J=10.5 Hz, H-8), 6.26 (dd, 1H, J=15.0, 10.5 Hz, H-7); ¹³C-NMR (125 MHz, CDCl₃) δ 12.81, 16.53, 26.90, 31.57, 32.82, 35.35, 37.65, 56.54, 79.90, 116.94, 124.16, 124.78, 126.84,

129.91, 131.84, 134.75, 136.33; FABMS m/z: 248 (M)⁺; HRFABMS calcd. for $C_{17}H_{28}O$ (M)⁺ 248.2140, found 248.2137.

(4R)-N-(tert-Butyloxy)carbonyl-2, 2-dimethyl-4-[(1Z, 5E, 7E, 11R)-11-methoxy-8-methyltetradeca-1, 5, 7, 13-tetraen-1-yl]thiazolidine (29).

A solution of 25 (602.1 mg, 0.986 mmol) in dry THF (12 mL) was cooled to -78 °C and LiHMDS (887 μL, 1.0 M in THF, 0.887 mmol) was added dropwise to the solution. The reaction mixture was stirred for 0.5 h at -78 °C. A solution of 4 (241.9 mg, 0.986 mmol) in dry THF (1 mL) was added dropwise to the resulting red solution at -78 °C over 20 min. The reaction mixture was allowed to warm slowly to 0 °C for 2 h and quenched by the addition of saturated aqueous NH_aCl (15 mL). The resulting mixture was extracted with EtOAc (3x24 mL), and the combined organic phases were washed with saturated aqueous NaCl (15 mL), dried and concentrated. Chromatography of the residue on silica gel (2 times, 120 g and 14 g, CH,Cl,/hexane 10/1 to CH₂Cl₂) gave **29** (264.2 mg, 60% yield) as a colorless oil, with recovery of **4** (60.9 mg, 25%): $[\alpha]_{0}^{25}$ +55.2° (c 0.99, CHCl₃); IR (CHCl₃) 2980, 2930, 1680, 1450, 1360, 1280, 1165, 1080, 970, 920, 850 cm⁻¹; ¹H-NMR (500 MHz, CDCl₃) δ 1.45 (s, 9H, (CH₃)₂C), 1.59 (td, 2H, J=8.0, 6.0 Hz, H-10'), 1.73 (s, 3H, CH₃-8'), 1.77 and 1.79 (s, 6H, (CH₂)₂-2), 2.01-2.33 (m, 8H, H-3', H-4', H-9' and H-12'), 2.57 (dd, 1H, J=12.0, 2.5 Hz, H-5), 3.19 (q, 1H, J=6.0 Hz, H-11'), 3.24 (dd, 1H, J=12.0, 6.5 Hz, H-5), 3.34 (s, 3H, OCH₃-11'), 5.04-5.12 (m, 3H, H-4 and H-14'), 5.43 (dt, 1H, J=10.5, 7.0 Hz, H-2'), 5.56 (dt, 1H, J=15.0, 7.0 Hz, H-5'), 5.69 (dd, 1H, J=10.5, 9.0 Hz, H-1'), 5.79 (d, 1H, J=10.5 Hz, H-7'), 5.81 (ddt, 1H, J=17.0, 10.0, 7.0 Hz, H-13'), 6.25 (dd, 1H, J=15.0, 10.5 Hz, H-6'); ¹³C-NMR (125 MHz, CDCl₃) δ 16.51, 27.59, 28.48 (x3), 29.63, 30.12, 31.55, 32.80, 34.25, 35.32, 37.59, 56.50, 61.01, 70.21, 79.81, 79.88, 116.91, 124.61, 127.24, 129.49, 131.23, 131.51, 134.68, 136.58, 152.35; FABMS m/z: 450 (M)⁺; HRFABMS calcd. for $C_{16}H_{44}O_3NS$ (M+H)⁺ 450.3042, found 450.2975.; Anal. Calcd for $C_{26}H_{43}O_3NS$: C, 69.44; H, 9.64; N, 3.11; S, 7.13. Found: C, 69.36; H, 9.71; N, 3.10; S, 6.99.

Curacin A (1).

TFA (970 µL, 12.6 mmol) was added to a solution of 29 (377.3 mg, 0.839 mmol) in water-saturated CH,Cl, (94 mL). The reaction mixture was stirred for 6 h and quenched by the addition of H₂O (20 mL). The organic phase was washed repeatedly with H₂O (20 mL), dried and concentrated. Chromatography of the residue on silica gel (38 g, 10/1 hexane/EtOAc) gave 30 (121.3 mg, 35% yield) as a pale yellow oil, with recovery of 29 (131.4 mg, 35%). The same procedure with the recovered 29 was repeated two more times to afford 30 (184.9 mg) in 54% total yield from 29. Bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BOPCl, 90.5 mg, 0.356 mmol) was added to a solution of 30 (121.3 mg, 0.296 mmol), (-)-(1R, 2S)-2methylcyclopropanecarboxylic acid (-)-7 (32.6 mg, 0.326 mmol) and Et₃N (103 μL, 0.739 mmol) in dry CH,Cl₂ (2.5 mL). The reaction mixture was stirred for 3 h, quenched by the addition of aqueous HCl (5%, 10 mL) and diluted with CH₂Cl₂ (20 mL). The organic phase was washed with saturated aqueous NaHCO₃ and NaCl (10 mL), dried and concentrated. Chromatography of the residue on silica gel (12 g, 15/1 hexane/EtOAc) gave the corresponding thiol ester (120.0 mg, 82% yield) as a pale yellow oil. TFA (376 µL, 4.88 mmol) was added to a solution of the foregoing thiol ester (120.0 mg, 0.244 mmol) in CH₂Cl₂ (3 mL) at 0 °C. The reaction mixture was allowed to warm to 20 °C for 2 h and concentrated under reduced pressure. Dry benzene (6 mL) was added to the resulting mixture and concentrated twice for replacing the reaction solvent with benzene. A solution of the residue containing the corresponding thiol ester ammonium salt in dry benzene (6 mL) was heated to reflux for 2.5 h and quenched by the addition of saturated aqueous NaHCO₃ (12 mL). The organic phase was washed with saturated aqueous NaCl (12 mL), dried and concentrated. Chromatography of the residue on silica gel (2 times, 12 g and 8 g, 15/1 hexane/EtOAc and 30/1 benzene/EtOAc) gave curacin A (1) (19.6 mg, 10% yield from 29) as a colorless oil: $[\alpha]_D^{25}$ +63.0° (c 0.77 CHCl₃), lit. $[\alpha]_D^{20}$ +62.0° (c 1.10, CHCl₃)⁶; IR (CHCl₃) 2930, 2860, 1720, 1640, 1610, 1430, 1360, 1280, 1170, 1140, 1070, 1000, 960, 910 cm⁻¹; The numbering of curacin A follows that reported by Gerwick et al.³ H-NMR (500 MHz, C_2D_4) δ 0.72 (td, 1H, J=8.5, 4.5 Hz, H-20), 0.94 (ddq, 1H, J=14.8, 8.5, 6.0 Hz, H-21), 1.18 (d, 3H, J=6.0 Hz, CH₃-21), 1.18 (m, 1H, H-20), 1.60 (m, 1H, H-19), 1.66 (m, 2H, H-12), 1.67 (s, 3H, CH₃-10), 2.00-2.27 (m, 8H, H-5, H-6, H-11 and H-14), 2.75 (dd, 1H, J=11.0, 10.0 Hz, H-1), 3.05 (dd, 1H, J=11.0, 8.5 Hz, H-1), 3.05 (m, 1H, H-13), 3.14 (s, 3H, OCH₃-13), 5.02-5.08 (m, 3H, H-2 and H-16), 5.41 (m, 1H, H-4), 5.54 (dt, 1H, J=15.0, 6.5 Hz, H-7), 5.65 (dd, 1H, J=10.5, 9.0 Hz, H-3), 5.83 (ddt, 1H, J=17.0, 10.0, 7.0 Hz, H-15), 5.97 (d, 1H, J=10.5 Hz, H-9), 6.34 (dd, 1H, J=15.0, 10.5 Hz, H-8); 13 C-NMR (125 MHz, C_6D_6) δ 12.33, 14.23, 16.00, 16.59, 20.11, 28.14, 32.17, 33.13, 35.79, 38.04, 39.95, 56.31, 74.34, 79.93, 116.79, 125.53, 127.88, 130.85, 131.31, 131.35, 135.32, 136.44, 168.40; FABMS m/z: 374 (M+H)*; HRFABMS calcd. for $C_{23}H_{16}$ ONS (M+H)* 374.2518, found 374.2524.

Preparation of the compound 27, 28 and (-)-7 was reported in detail in our previous paper. 14

References and notes

- For the details on the structures and functions of microtubules, see the following review articles. (a)
 Dustin, P., Microtubules, Springer-Verlag, New york, 1978. (b) Soifen, D., Ed., Dynamic Aspects of
 Microtubule Biology, New York Academy of Science, New York, 1986. (c) Avila, J., Ed., Microtubule
 Proteins, CRC, Boca Raton, FL, 1990. (d) Hamel, E., in Microtubule Proteins, Avila, J., Ed., CRC,
 Boca Raton, FL, 1990.
- 2. Iwasaki, S. Med. Res. Rev. 1993, 13, 183-198.
- Gerwick, W. H.; Proteau, P. J.; Nagle, D. G.; Hamel, E.; Blokhin, A.; Slate, D. L. J. Org. Chem. 1994, 59, 1243-1245.
- Nagle, D. G.; Geralds, R. S.; Yoo, H. -D.; Gerwick, W. H.; Kim, T. -S.; Nambu, M.; White, J. D. Tetrahedron Lett. 1995, 36, 1189-1192.
- 5. Onoda, T.; Shirai, R.; Koiso, Y.; Iwasaki, S. Tetrahedron Lett. 1995, 36, 5765-5768.
- (a) White, J. D.; Kim, T. -S.; Nambu, M. J. Am. Chem. Soc. 1995, 117, 5612-5613.
 (b) Hoemann, M. Z.; Agrios, K. A.; Aubé, J. Tetrahedron Lett. 1996, 37, 953-956.
 (c) Ito, H.; Imai, N.; Tanikawa, S.; Kobayashi, S. Tetrahedron Lett. 1996, 37, 1795-1798.
 Ito, H.; Imai, N.; Takao, K.; Kobayashi, S. Tetrahedron Lett. 1996, 37, 1799-1800.
 (d) Onoda, T.; Shirai, R.; Koiso, Y.; Iwasaki, S. Tetrahedron Lett. 1996, 37, 4397-4400.
- 7. Fukuyama, T.; Xu, L. J. Am. Chem. Soc. 1993, 115, 8449-8450.
- 8. Kemp, D. S.; Carey, R. I. J. Org. Chem. 1989, 54, 3640-3646.
- 9. Duthaler, R. O. Angew. Chem. Int. Ed. Engl. 1991, 30, 705-707 and personal communication.
- 10. Bergman, R. G. J. Am. Chem. Soc. 1969, 91, 7405-7411.
- 11. Cabré, J.; Palomo, A. L. Synthesis 1984, 413-417.
- 12. Takahashi, M.; Iwasaki, S.; Kobayashi, H.; Okuda, S.; Murai, T.; Sato, Y.; Haraguchi-Hiraoka, T.; Nagano, H. J. Antibiotics 1987, 40, 66-72.
- 13. Hafner, A.; Duthaler, R. O.; Marti, R.; Rihs, G.; Rothe-Streit, P.; Schwarzenbach, F. J. Am. Chem. Soc. 1992, 114, 2321-2336.
- 14. Onoda, T.; Kawai, N.; Shirai, R.; Iwasaki, S. Tetrahedron in press.
- (a) Yonetani, K.; Hirotsu, T.; Shiba, T. Bull. Chem. Soc. Jpn. 1975, 48, 3302-3305.
 (b) Hirotsu, Y.; Shiba, T.; Kaneko, T. Bull. Chem. Soc. Jpn. 1967, 40, 2945-2949.
- 16. Cicala, G.; Curci, R.; Fiorentino, M.; Laricchiuta, O. J. Org. Chem. 1982, 47, 2670-2673.
- 17. Kocienski, P. J.; Cernigliaro, G.; Feldstein, G. J. Org. Chem. 1977, 42, 353-355.
- 18. Yao, Z.-J.; Wu, Y.-L. J. Org. Chem. 1995, 60, 1170-1176.
- 19. Bergelson, L. D.; Barsukov, L. I.; Shemyakin, M. M. Tetrahedron 1967, 23, 2709-2720.
- 20. Takeda, K.; Kobayashi, T.; Saito, K.; Yoshii, E. J. Org. Chem. 1988, 53, 1092-1095.
- 21. Yan, L.; Kahne, D. Synlett 1995, 523-524.
- (a) Tsuji, T.; Kataoka, T.; Yoshioka, M.; Sendo, Y.; Nishitani, Y.; Hirai, S.; Maeda, T.; Nagata, W. Tetrahedron Lett. 1979, 30, 2793-2796. (b) Fuji, K.; Ichikawa, K.; Node, M.; Fujita, E. J. Org. Chem. 1979, 44, 1661-1664. See also, (c) Akiyama, T.; Shima, H.; Ozaki, S. Synlett 1992, 415-416. (d) Akiyama, T.; Hirofuji, H.; Ozaki, S. Tetrahedron Lett. 1991, 32, 1321-1324.
- 23. Onoda, T.; Shirai, R.; Iwasaki, S. unpublished results.